# Safety and protective effect of *Lactobacillus acidophilus* used as probiotic agent in goats

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### **Abstract**

The study was aimed to study the protective effect of *lactobacillus acidophilus* isolated from fresh sheep milk. the result of this study was revealed that Lactobacillus isolates had liver improvement functions. Lactate & Sorbitol dehydrogenase activities of goats dosed Lactobacillus isolates alone were lower than the control. There were reduction in the count of enterobactria in goat dosed with Lactobacillus after three days. Protection of gastrointestinal tract by these isolates was also observed.

#### Introduction

Gastrointestinal disorders are caused factors including various antibiotic by administration (29) or as a result of infectious agents such toxogenic as Escherichia coli., Salmonella enteritidis . Entamoeba histolotica viruses (27). Innovative approaches have been tried as alternative to antibiotics in treating gastrointestinal diseases and these include using live biotherapeutic agents such as yeast (Sacchromycess Spp) and bacterial isolates (Lactobacillus Spp). Or fecal anemals (14).Lactobacillus are important for maintenance of the intestinal microbial ecosystem (26). Colonization of the gut with lactobacilli start within the first week of life (25). The presence of this group of bacteria in the gut is considered to have several potential benefits such as growth promotor of farm animals (2), protection against pathogens (7), alleviation of lactose intolerance (17), relief of constipation (30), anticholesterolemic effect (5) and

immunomodulation (1).Lactobacilli exert protective therapeutic their effect through production of antimicrobial compounds (10,21), reduction of gut pH by stimulating the acid producing lactic microflora (11), competition with binding receptor sties that pathogens occupy (20,22), stimulating of immunomodulatory cells (24) and competition with pathogens for a viable nutrients (11,24). Walker and suggested Duffy that current (30)perspectives on biotechnological applications of probiotic products require further in vitro and in vivo investigation to evaluate the safety of using wild type organisms or those obtained by genetic engineering. The present study is therefore aimed for understanding the protective effect of Lactobacillus acidophilus from fresh sheep milk and their ability to reduce the toxologic and pathologic consequence with enterotoxogenic associated used to experimentally infected goats.

#### **Materials and Methods**

Lactobacillus acidophilus were isolated from fresh sheep milk on MRS agar. The isolates were characterized using colonial morphology and biochemical tests according to (6). These Lactobacillus spp were also found to adhere to the ilial epithelial cells of goats. The isolates were inoculated in MRS broth and incubated at 37 C° for 2 days to obtain large concentration about 10<sup>10</sup> CFU/ ml.These cells were washed. suspended lyophilized rehydrated skim milk and

stored at -20 C° until use (13). The concentration of a viable cells was determined by serial dilution techniques (28). Twenty four (24) goats were used in this study they were randomly assigned to 4 treatment groups each was made up of 6 goats per groups. Lyophilized lactobacillus cells were reconstituted by dissolving 1 gr in 10ml of normal saline (approximatly 10<sup>10</sup> CFU/ml). The first group was kept on basal diet alone and considered as control group. Second group fed on the basal diet

and were also dosed with 0.3ml of L.acidophilus. Third group were fed on the basal diet , dosed with 0.3 ml of L.acidophilus and infected with 0.3 ml of 10<sup>5</sup> CFU/ml of enterotoxogenic *E.coli*. fourth group was fed on basal diet and infected with 0.3 ml of 105 CFU/ml of enterotoxogenic E.coli. The treatment above was repeated on the second day. A post ingestion period of 18 day was observed after administration of culture and blood sample were collected for serum biochemical analysis for the following parameter using kits from Boehringer-Mannheim Company – Germany.

- 1- Sorbitol dehydrogenase
- 2- Lactate dehydrogenase
- 3- Aspartate amino transferees

- 4- Alkaline phosphates
- 5- Cholesterol

Freshy voided fecal materials were collected and pooled from each goat (1gr/goat) at days zero and 3. The faeces were homogenized in normal saline and serially diluted and plated on MRS agar for the enumeration of lactobacilli and MacConkey's agar for enumeration entrobacteriacea especially *E.coli*. plates were incubated at 37 C° for 24 hours and colony forming unites on the plates were recorded (19). The data gathered from toxocologic assay and faecal flora were processed using one way analysis of variance (ANOVA).SPSS. 12. The level of significance was set at (P<0.05). Means were compared by Duncan test.

#### **Result and Discussion**

The aspartate amino transferase (AST) activity in animals of third group was highest and significantly different (P<0.05) from the control (First group) (Table 1). AST is an enzyme that increased in activity in diseases such as severe bacterial infection and tumors of organ such as heart and muscle (8).Lactobacilli can translocate and survive in the spleen, liver and lungs (4,6). In the course of their translocation they can cause cellular injury that may increase AST level in the serum. This may account for increase in AST observed for goats of third group compared with the control group. The higher AST level in third group may be due to the combine activities of Lactobacillus and E.coli in the GIT. In their study (27) reported that, to obtain protective effect in animals, treatment with probiotic agents had to be initiated 10 days before challenge with pathogens but in this report oral dosing with Lactobacillus and challenge with E.coli was simultaneous. The results of lactate and sorbitol dehydrogenases (LDH and SDH) activities in the serum revealed that., The third group were higher (P<0.05) significantly than control. LDH and SDH is principally found in the liver and together is regarded as being more specific than AST alone for detecting liver cell damaged (8,18). The

implication of this result is that, there is a pronounced toxocologic effect in goats of the third and fourth groups. The lower LDH and SDH in goats treated with lactobacillus alone (Second compared with the control indicate liver function improvement brought about by the lactobacillus. Hepatocytes play a major role in absorbing and metabolizing many toxic chemicals (12). They are therefore liable to injury by various chemicals including food. The alkaline phosphatase (ALP) activity of goats treated with lactobacillus and E.coli (Third group) and those treated with E.coli alone (fourth group) were significantly higher than those treated with lactobacillus alone (Second group). A rise of ALP activity has been linked with an increased osteoblastic of bile flow activity (3) and lack (Cholestasis). Only slight cholesterolemic effect was also observed in goats treated with lactobacillus. Lactobacilli has been found to have direct effect on cholesterol level by assimilation and removal from the growth medium. This has been demonstrated in pigs (15) and rats (5). Serum ALP levels has been reported to increase with increase in the serum cholesterol (18).The ability isolates to protect the GIT pathogens can be confirmed by monitoring

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the count of enterobacteria especially E.coli and beneficial bacteria especially lactobacilli in goats faeces (7). There was an increase in fecal lactobacilli count in goats treated with lactobacilli (Table2).A slight decrease in enteric bacteria count was also observed in most of the goats. There was increase enterobacteria and lactobacilli count from day zero to day 3 in both control groups (First and fourth groups). In a similar study (9) reported an increase of lactobacilli count in faeces of rats that was basal diet devoid of probiotic agents. The high lactobacilli count in goats

treated with lactobacilli and E. coli (Third group) may be responsible for the partial protection of the GIT of goats in this group. Earlier report showed that a selected probiotic strain L. reuteri and L. acidophilus showed an increasing effect in numbers of enterobacteria in piglets (23). The ability of lactobacilli to produce toxic metabolites such as lactic acid, hydrogen peroxide  $(H_2O_2)$ and bacteriocins has been suggested as being responsible for their ability to inhibit other bacteria (19). Other factors such as host immunomodulation also play a prominent role. (13).

Table (1): Serum biochemical markers in different groups

Group	AST	LDH	SDH	ALP	Cholesterol
	/IU/L	/IU/L	/IU/L	/IU/L	Mg/d/
1 St group	182.612 ± 32.3 a	128.47 ±	19.43 ±	102.9 ±	93.5 ±
		22.91 a	2.01 a	1.223 a	7.216 a
2nd group	233.623 ±	80.32 ±	14. 26 ±	104.263 ±	29.63 ±
	12.621 b	7.81 b	1.75 a	1.26 a	6.22 b
3ed group	635.264 ± 33.3 c	520.83 ±	68. 67 ±	1560.12 ±	165.12 ±
		22.6 c	14.22 b	31.12 b	3.61 c
4th group	422. 621 ±	160.4 ±	45.02 ±	520.62 ±	92. 63 ±
	12.25 d	11.3 d	4.22 c	11.12 c	6.22 a

Table (2): Total count of faecal bacteria  $\times 10^6$ /ml

Group	Enterobaci	teriaceae	Lactobacillus		
Group	Day zero	Day 3	Day zero	Day 3	
1 St group	5.72 ±	6.61 ±	6.12 ±	7.22 ±	
1 St group	0.82 a	1.23 b	0.51 a	0.21 b	
2nd group	5.52 ±	5.32 ±	5.32 ±	8.36 ±	
zna group	0.863 a	0.36 b	0.621 a	0.261 b	
2 ad amoun	5.92 ±	5.71 ±	5.39 ±	$7.38 \pm$	
3ed group	0.29 a	0.42 b	0.52 a	0.76 b	
Ath group	5.56 ±	6.99 ±	5.22 ±	6.21 ±	
4th group	0.36 a	0.42 b	0.59 a	0.30 b	

odifferent litters means significant differences.

#### References

- 1. Aottouri, N.; Bouras, M; Tome, D; Marcos .A: Lemonnier, D (2002). Ingestion of lactic acid bacteria by rats increases lymphocytic proliferation and interferon gama production. Br. J. Nut, 87:367-373.
- 2.Baird, D.M (1977) probiotics help boost efficiency. Feed stuffs. 49:11-12.
- 3.Baron, D.N.; Whicher, J.T. and Lee, K.E. (1994). A new short textbook of chemical pathology, 5<sup>th</sup> edition. ELBS. Pp:151-156.

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- 4.Berg, R.D. (1983).Translocation of indigenous bacteria from the intestinal In tract. human intestinal microflora in health and disease. Henteges. D.J. London. Academic press Pp: 333-352.
- 5. Bertazzoni, M.E.; Benini, A; Marzotto, M; Hendriks, H. Sbarbati, A and Dellaglio, F.(2001). **Preliminary** screening of health promoting properties of new lactobacillus strains in vitro and in vivo. HEALFO abstracts. Italy.
- 6.Bloskma, N.; Ettekoven, H; Hothious, F.M; Van Noorle - Jansen , L; De- Reuver, M.J; Krwwflenberg, J.G. and Willers, J.M. (1981). Effects lactobacilli of parameters of non specific resistance of mice. Med. Microbiol. Immunol. 170: 45-53.
- 7. Casas, I.A. and Dobrogosz, W.J (2000). Validation of probitic concept Lactobacillus reuteri confers broad spectrum protection against disease in human and animals. Microbial. Ecol. Health Dis. 12:247-285.
- 8. Cheesborough, M (1991).Medical laboratory manual for tropical  $2^{\rm nd}$ countries edition. **Topical** health Technology and Scientific limited. Butterworth (1): 494-529.
- 9.Chung, H.G.(2003). Control of food bone Pathogens by bacteriocin like substance from Lactobacillus Spp in combination with high pressure processing. PhD thesis the Ohio university .
- 10. Dood, H.M. and Gasson, M.J. (1994). Bacteriocins of lactic lcid bacteria. In Gassons, M.J. and de Vos, W.M (Eds) Genetics and biotechnology of lactic acid bacteria. Glassgow. United Kingdom: Blackie Academic and Professional: 211-251.
- 11. Edens, F.W. (2003). An alternative for antibiotic use in poultry: Probiotic. Rev. Bras. Sci. Avic ; 5(2):101-134.

- 12. Eka, O.U; Zagi, M.M; and Umoh, I.B (1994). Toxicologic studies monosodium glutamate Α review. Biochemistry (4):57-74.
- 13. Fujiwara, S; Seto; Y; Kimura, A and Hashiba, H (2001) Establishment of orally administrated Lactobacillus gasseri SBT 2055 SR in the gastrointestinal tract of human and its influence intestinal microflora and metabolism. J. Appl. Microbiol. 90:343-352.
- 14. Fuller. R. (1992).Probiotic. The Scientific basis (Ed). London. Champan and Hall.
- 15. Gilliland, S.E. Nilson, C.R. Maxwell, C (1985). Assimilation of cholesterol by Lactobacillus acidophilus. Appl. Environ. Microbiol. 49: 37-381.
- 16. Hatcher, G.E. and Lambercht (1993). Augmentation of macrophage phagocytic activity by cell free extracts of selected lactic acid bacteria. J. Dairy Sci. 76:2485-2492.
- 17. Jiang, T; Mustapha, A and Savaiano, D.A (1996). Improvement of lactose digestion in human by ingestion of unfermented milk containing *Bifidobacterium* longum. J. Diary. Sci. 79(5): 750-757.
- D.E. (1999).18. Johnston, Special consideration in interpreting liver function The American test. family Academy of Physician. April 15,1999.
- 19. Juven, B.J.; Schved, F and Linder, P (1992). Antagonistic compounds by chicken intestinal produced strain of Lactobacillus acidophilus. J. Food Prot. 55:157-161.
- 20. Kailasapathy, K. and Chin, J (2000). Survival and therapeutic potential of probiotic organism with reference of Lactobacillus acidophilus and Bifidobacterium Imminol. Spp. Cell Biol. 78: 80-88.

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- 21. Kalenhammer, T.R. (1993). Genetic of bacteriocins produced by lactic acid bacteria. FEMS. Microbiol. Rev; 12:39-86.
- 22. Ohashi, N; Inoue, R; Tanaka, K; Usema, Y and Ushida, K. (2002). Strain gauge force tranducer and its application in a pig model to the effect of probiotics on colonic motility. J. Nut & Vitaminology (Tokyo); 47(5): 351-356.
- 23. Ratcliff, B; Cole, C.B; Fuller, R and Newport. M.J (1958). The effect of yoghurt and fermented milk with porcine intestinal strains of L. rueteri on the performance gastrointestinal infection. Microbiol. 3:203-211.
- 24. Rolfe , R.D. (2000). The role of probiotic culture in the control of gastro intestinal heath. J. Nut. 130 (25):3965-4025.
- 25. Salminen. S; Isolouri, E and onnella, T (1995). Gut flora in normal and disordered states. Chemotherapy. 41:5-15.

- 26. Sandine, W.E (1979).Role of lactobacillus in the intestinal tract. J. Food Protect. 42: 259-262.
- 27. Sliva, A.M; Bambirra , E.A; Oliveira, A.L; Souza, P.P; Gomes. D.A; Vieira, E.C and Nicoli , J.R Protective (1999).effect bifidus milk on the experimental infection with Salmonella enteritidis in mice. J. Appl. Microbiol. 86:331-336.
- 28. Tylor, J. (1962). The estimation of bacterial numbers by ten fold dilution series. J. Appl. Bacteriol . 25:54-61.
- 29. Van der Waaij, D; Horstra, H and Wiegersma, N (1982). Effect of antibiotic in lactum resistance of the digestive tract to colonization. J. Infect. Dic. 146: 417-422.
- 30. Walker, A.W. and Duffy, L.C (1998). Diet and bacterial colonization. Role of probiotics and Prebiotics : Review. J. Nutr. Biochem. 9:668-675.

# التأثير الوقائي والأمين للعصيات اللبنية المحبة للحموضة المستخدمة كمعزز حيوى في الماعز

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## الخلاصة

المعزولة المعروفة التأثير الوقائي للعصيات اللبنية المحبة للحموضة Lactobacillus acidophilus المعزولة من حليب الأغنام الطازج كمعزز حيوي Probiotic في الماعز. وقد أظهرت نتائج الدراسة ان هذه الجراثيم أدت الي تحسين وظائف الكبد حيث أظهرت فعالية إنزيمي اللاكتيك ديهايدروجينيز والسوربيتول ديهايدروجينيز فعالية إنزيمي اللاكتيك ديهايدروجينيز والسوربيتول ديهايدروجينيز Sorbitol dehydrogenase انخفاضا في مجموعة الحيوانات التي جرعت بالعصيات اللبنية المحبة للحموضة فقط مقارنة مع مجموعة السيطرة كما أظهرت الدراسة حصول اختزال في عدد الجراثيم المعوية في الحيوانات التي جرعت العصيات اللبنية يعد ثلاثة أيام من التجريع وقد تمت ملاحظة التأثير الواقى لهذه الجراثيم على القناة الهضمية .