

## Effect of Dietary Vegetable Wastes as Prebiotics on Serum Biochemical Parameters and Intestinal Microflora in Juveniles of the Common Carp *Cyprinus carpio*

Mustafa A.A. Albadran<sup>1</sup>, Salah M. Najim<sup>1</sup> & Khalidah S. Al-Niaeem<sup>2\*</sup>

<sup>1</sup>Department of Fisheries and Marine Resources, <sup>2</sup>Unit of Aquaculture, College of Agriculture, University of Basrah, Basrah, Iraq

\*Corresponding author: kalidah\_salim@yahoo.com

**Abstract:** Effects of replacing wheat flour in fish feed with vegetable wastes on some biochemical parameters and normal flora of juvenile common carp *Cyprinus carpio* were studied in laboratory. Three replacing ratio (5, 7.5 and 10%) were investigated. The total protein and albumin concentration in blood serum increased significantly ( $p < 0.05$ ) in all feeding treatments compared with control treatment. Highest concentrations were achieved by fish of feed group T3 (94.63 and 5.72 mg/100 ml respectively). Levels of total cholesterol (TC), triglycerides (TG), high-density lipoprotein and low-density lipoprotein (LDL) were decreased significantly ( $p < 0.05$ ) in fish with 10% replacing ratio (being 130.86, 46.10, 85.46, 3.26 and 9.22 mg/100 ml, respectively) compared with control diet. Population of the lactic acid bacteria (intestinal microflora) increased significantly ( $p < 0.05$ ) after feeding experiment in all treatments and reached the highest in T3 (2.93 Log CFU/g). The results reveal that supplementation of dietary vegetable wastes have most positive effects on improving the beneficial intestinal microbiota and on some haemato-serological parameters of juveniles of *C. carpio*. Future studies are needed to focus on disease challenge and stress response as vegetable wastes have been known as a good prebiotic candidate for applications with common carps.

**Keywords:** Fishes, Prebiotics, Vegetable Wastes, Serum Biochemistry, Intestinal Microflora.

### Introduction

During the last few decades, the intensification of aquaculture production, diseases and deterioration of environmental conditions are major problems in fish farming and face massive economic losses in prevention and control of diseases, antibiotics used as traditional strategy and also in fish growth as well as efficacy of feed conversion (Ahmadifar et al., 2011). Development of antimicrobial resistant pathogens were recognized but there is a huge risk of transmission of resistance bacteria from aquatic environment to humans (Ringø et al., 2014).

Using antibiotics is harmful for aquatic animals as they kill beneficial microbiota in their gastrointestinal system and also they accumulate in fish

products which become unsafe for human consumption. Considering these factors, the improvements of non-antibiotic agents are more suitable for health management in aquacultures (Ringø et al., 2010a).

Dietary supplement such as prebiotics provide nonspecific disease protection and also act as growth promoting factors. The use of prebiotics, nondigestible dietary ingredients beneficially affects the host by selectively stimulating the growth of and/or activating the metabolism of health-promoting bacteria in the intestinal tract is a novel concept in aquaculture (Momeni-Moghaddam et al., 2015; Forsatkar et al., 2017). The dietary administration of prebiotics can exert some effect on the intestinal microbiota, digestive enzyme activities, morphology and volatile short chain fatty acids (VSCFAs) production in various aquatic animal species (Ringø et al., 2014).

This study was carried out to investigate the effect of replacing ratio (feed diets) of vegetable wastes on biochemical condition of blood serum and normal flora in common carp juveniles with different concentrations of replacements.

## Materials and Methods

The vegetable wastes of lettuce (*Lactuca sativa*), green onion (*Allium cepa*), garlic (*Allium sativum*) and kale (*Brassica oleracea*) were collected from a local market in Basrah city, Southern Iraq. All vegetable wastes were dried in room temperature for 72 hours and ground by using electric grinder. Ground material was sieved through a 0.4 mm mesh for homogenization and kept in marked plastic bags.

Juveniles of the Common carp were obtained from the University of Basrah fish farm. A total of 144 specimens with initial mean weights of  $13.71 \pm 0.57$  g were randomly distributed among 12 aquarium tanks (12 fish per tank, three replicates per treatment). Water temperature, dissolved oxygen, pH and salinity (‰) were measured before and after each experiment.

The experimental diets used in fish feed were manufactured by the Marine Science Center, University of Basrah. The diets were formulated from fishmeal, soybean meal, corn oil, barley flour and wheat flour. The feed replacement which included vegetable wastes was added at percentage of 5%, 7.5% and 10% replacing wheat flour in fish diets (Table 1). Fishes were fed 5% body weight per day. Proximate composition of the experimental diet is shown in Table (2).

Blood samples from each fish in the different groups were collected by suction of the caudal peduncle. Blood serum was then separated by centrifugation for three minutes at 1500 rpm (Yang & Chen, 2003). The levels of serum biochemical parameters were assayed according to the instructions provided with the corresponding biochemical kits. These

parameters were total protein (TP), albumin (AL), total cholesterol (TC), triglycerides (TG) and high density lipoprotein (HDL).

Globulin concentration was calculated directly according to Wolf & Darlington (1971) as:

$$\text{Globulin} = \text{Total protein} - \text{albumin}$$

Low density lipoprotein (LDL) cholesterol concentration was calculated according to Taherpour et al. (2009) as:

$$\text{LDL} = \text{Total cholesterol} - (\text{HDL} + \text{VLDL})$$

One-way analysis of variance (ANOVA) was made by using SPSS software (version 18) and the difference between the means and the significant differences was verified by using RLSD test.

The bacterial counts (TBC) and lactic acid bacteria (LAB) were assayed according to Andrews et al. (1992).

Table 1. Fish experimental diet composition (%).

Ingredients	Feed group			
	Control (T1)	5% (T2)	7.5% (T3)	10% (T4)
Fishmeal	20	20	20	20
Soybean meal	15	15	15	15
Barely flour	20	20	20	20
Corn oil	5	5	5	5
Wheat flour	40	35	32.5	30
Vegetable wastes	0	5	7.5	10

Table 2. Proximate composition of the experimental diet.

Ingredients	Feed group			
	Diet (% dry weight)			
	Control (T1)	5% (T2)	7.5% (T3)	10% (T4)
Protein	32.56	31.8	30.84	31.24
Lipid	9.11	9.65	10.02	9.23
Carbohydrates	49.52	49.36	50.31	50.68
Moisture	2.88	3.17	3.24	2.96
Ash	5.93	6.02	5.59	5.89

## Results

Values of water quality parameters are shown in Table (3). They were adequately stable; temperature (27.1-27.7 C°), pH (8.8-8.9), dissolved oxygen (8.8-9.4 mg/l) and salinity (2.1-2.3‰).

Table 3. Water quality parameters of rearing water for common carp juvenile during the experiment.

Parameter	Replacing ratio (%)			
	feed group (Mean $\pm$ S.D)			
	Control (T1)	5% (T2)	7.5% (T3)	10% (T4)
Temperature (C <sup>0</sup> )	27.6 $\pm$ 0.24	27.1 $\pm$ 0.24	27.7 $\pm$ 0.30	27.7 $\pm$ 0.20
pH	8.85 $\pm$ 0.02	8.8 $\pm$ 0.03	8.9 $\pm$ 0.04	8.9 $\pm$ 0.04
Dissolved oxygen (mg/l)	8.8 $\pm$ 0.2	8.8 $\pm$ 0.4	9.4 $\pm$ 0.2	9.4 $\pm$ 0.2
Salinity (‰)	2.3 $\pm$ 0.20	2.3 $\pm$ 0.10	2.2 $\pm$ 0.20	2.1 $\pm$ 0.04

The general biochemical indices of juveniles of the common carp before and after feeding on experimental feeds which contain different inclusion replacing ratios are presented in Table (4).

Table 4. Blood serum biochemical (mg/100 ml) of common carp juvenile before and after feeding on experimental feeds.

Parameter mg/100 ml	Replacing ratio (%)			
	feed group (Mean $\pm$ S.D)			
	Control (T1)	5% (T2)	7.5% (T3)	10% (T4)
Total protein	3.86 $\pm$ 0.58 <sup>a</sup>	4.39 $\pm$ 0.67 <sup>a</sup>	4.68 $\pm$ 0.06 <sup>b</sup>	5.72 $\pm$ 0.75 <sup>c</sup>
Albumin	2.89 $\pm$ 0.45 <sup>a</sup>	3.26 $\pm$ 0.67 <sup>b</sup>	3.70 $\pm$ 0.21 <sup>b</sup>	4.63 $\pm$ 0.12 <sup>c</sup>
Globulin	1.10 $\pm$ 0.13 <sup>a</sup>	1.13 $\pm$ 0.23 <sup>a</sup>	0.98 $\pm$ 0.27 <sup>a</sup>	1.08 $\pm$ 0.65 <sup>a</sup>
Total cholesterol	208.10 $\pm$ 30.11 <sup>a</sup>	189.30 $\pm$ 1.27 <sup>ab</sup>	169.96 $\pm$ 9.60 <sup>b</sup>	139.86 $\pm$ 4.3 <sup>c</sup>
Triglycerides	123.42 $\pm$ 20.86 <sup>a</sup>	87.25 $\pm$ 2.57 <sup>ab</sup>	84.95 $\pm$ 4.83 <sup>b</sup>	46.10 $\pm$ 5.26 <sup>c</sup>
HDL	110.34 $\pm$ 21.66 <sup>a</sup>	92.31 $\pm$ 7.10 <sup>ab</sup>	72.67 $\pm$ 5.96 <sup>bc</sup>	85.46 $\pm$ 2.23 <sup>c</sup>
LDL	91.51 $\pm$ 3.27 <sup>a</sup>	79.48 $\pm$ 2.03 <sup>b</sup>	67.17 $\pm$ 2.92 <sup>bc</sup>	63.26 $\pm$ 1.67 <sup>c</sup>

Values in the same raw with different superscript letters are significantly ( $p < 0.05$ ) different.

The total bacterial counts (TBC) and lactic acid bacterial counts (LAB) were significantly ( $P < 0.05$ ) increased by the addition of 10% and 7.5% replacing ratios of vegetable wastes than in control fishes (Table 5).

Table 5. Bacterial counts in the intestine of common carp juvenile during the experiment.

Intestinal Microflora (Log CFU/g)	Replacing ratio (%)			
	feed group (Mean $\pm$ S.D)			
	Control (T1)	5% (T2)	7.5% (T3)	10% (T4)
TBC	5.31 $\pm$ 0.05 <sup>b</sup>	5.53 $\pm$ 0.06 <sup>b</sup>	5.70 $\pm$ 0.06 <sup>c</sup>	5.87 $\pm$ 0.02 <sup>c</sup>
LAB	2.20 $\pm$ 0.17 <sup>a</sup>	2.41 $\pm$ 0.09 <sup>a</sup>	2.71 $\pm$ 0.05 <sup>b</sup>	2.93 $\pm$ 0.02 <sup>b</sup>

Values in the same raw which carry different superscript letters are significantly ( $p < 0.05$ ) different.

## Discussion

Prebiotics are functional dietary supplements with documented positive applications in a range of fish and crustacean species (Ringø et al. 2010a). They can change the composition of the normal flora by changing the types of substrates available to bacteria and make them more appropriate to the micro-ecophysiological conditions of the resident intestinal microflora which allow them to grow rapidly (Ringø et al. 2010b; Carbone & Faggio, 2016). Haematological and biochemical parameters are considered as useful measuring instruments in order to check the quality of fish health and physiological responses, nutrient absorption and animal surroundings that affect fishes (Hoseinifar et al., 2011)

Values of water quality parameters indicated in Table 1 are within the suitable ranges for culture of the common carp according to Marković et al. (2009).

The results of the present study show that supplementation with prebiotic derived from a vegetable wastes powder: fructo-oligosaccharides (FOS), mannan-oligosaccharides (MOS) and xylo-oligosaccharides (XOS) have positive effects on the bacterial populations in the intestine of the experimental fishes (Eldahshan & Singab, 2013; Gebreyohannes & Gebreyohannes, 2013; Mageney et al., 2017). The degree of the positive effects on the bacterial populations in the intestine was affected by the prebiotic dose (Ringø et al., 2010b; Forsatkar et al., 2017).

The results in Table (4) indicated that the total proteins, albumin and globulin increased. The total proteins in the serum include albumin and globulin. Globulin is made up of fractions of a1, a2, b and c globulins that made immunoglobulin (Ig), the increase in the levels of serum protein, albumin and globulins in fishes is thought to be associated with a stronger innate immunity response (Ringø et al., 2014).

The present study also showed that there were significant differences in the serum biochemical parameters such as cholesterol and triglyceride, between treatments (Table 4). Andrews et al. (2009) observed a significant improvement in fed MOS supplemented diet in comparison with those fed on the control diet. Ye et al. (2011) showed that dietary FOS and MOS decreased flounder cholesterol and triglyceride compared to the control. Ahmed (2014) and Abdulrahman et al. (2016) observed significant differences of different levels of FOS on some blood indices in young common carp.

Total bacterial count (Table 5) increased with increasing prebiotic doses. This indicates that the doses of prebiotics provided a substrate for the growth of bacteria in the intestine, which led to larger bacterial populations in the treated groups than in the control (Ye et al., 2011). In the present study, significant increase in lactic acid bacteria (LAB), especially in the 10%

replacing ratio was noticed. Lactic acid bacteria are able to produce bacteriocins, lactic acid and other antagonistic compounds, which inhibit the growth of certain fish pathogens and hence they improve the fish intestinal ecosystem (Ringø et al. 2006). They are not generally considered to be the dominant species in the gut, but prebiotics shift the normal flora to them and increase their population (Ringø et al. 2010a, b).

The results of the present study indicate that prebiotic supplementation (vegetable wastes) influenced fish biochemical parameters and normal flora. Such results are in agreement with earlier studies (Akrami et al., 2012; 2013, Hoseinifar et al., 2014 a, b; Amani et al., 2015; Hoseinifar et al., 2016).

## Conclusions

In conclusion, the results observed in this trial reveal that supplementation of dietary vegetable wastes appears to be most positive for beneficial intestinal microbiota and some biochemical parameters of juveniles of the common carp. It is necessary that future studies would be focused on disease challenge aspects and stress response as vegetable wastes are known as a good prebiotic candidate for applications with common carps.

## References

- Abdulrahman, N.M.; Ahmed, V.M.; Hama Ameen, H.J.H. & Hasan, B.R. (2016). Study the effect of different level of fructooligosaccharide (FOS) on some blood indices in young common carp (*Cyprinus carpio* L.). Basrah J. Vet. Res., 15 (3): 34-44.
- Ahmadifar, E.; Akrami, R.; Ghelichi, A.; Mohammadi Zarejabad, A. & (2011). Effects of different dietary prebiotic inulin levels on blood serum enzymes, hematologic and biochemical parameters of great sturgeon (*Huso huso*) juveniles. Comp. Clin. Pathol. 20 (5): 447-451.
- Ahmed, V.M. (2014). Comparative effects of probiotic (*Saccharomyces cerevisiae*), prebiotic (Fructooligosaccharide FOS) and their combination on growth performance and some blood indices in young common carp (*Cyprinus carpio* L.). M. Sc. Thesis, Fac. Agric. Sci., Univ. Sulaimani, 97 pp.
- Akrami, R.; Mansour, M.R.; Chitsaz, H.; Ziaei, R. & Ahmadi, Z. (2012). Effect of dietary mannan oligosaccharide on growth performance, survival, body composition and some hematological parameters of carp juvenile (*Cyprinus carpio*). J. Anim. Sci. Adv., 2 (11): 879-885.
- Akrami, R.; Razeghi Mansour, M.; Ghobadi, S.H.; Ahmadifar, E.; Shaker Khoshroudi, M. & Moghimi Haji, M.S. (2013). Effect of prebiotic mannan oligosaccharide on hematological and blood serum biochemical parameters of cultured juvenile great sturgeon (*Huso huso* Linnaeus, 1754). J. Appl. Ichthyol., 29 (6): 1214-1218.

- Amani Denji, K.; Razeghi Mansour, M.; Akrami, R.; Ghobadi, Sh.; Jafarpour, S.A. & Mirbeygi, S.K. (2015). Effect of dietary prebiotic mannan oligosaccharide (MOS) on growth performance, intestinal microflora, body composition, haematological and blood serum biochemical parameters of rainbow trout (*Oncorhynchus mykiss*) juveniles. *J. Fish. Aquat. Sci.*, 10 (4): 255-265.
- Andrews, S.R.; Sahu, N.P.; Pal, A.K. & Kumar, S. (2009). Haematological modulation and growth of *Labeo Rohita* fingerlings: Effect of dietary mannan oligosaccharide, yeast extract, protein hydrolysate and chlorella. *Aquac. Res.*, 41 (1): 61-69.
- Andrews, W. (1992). Manual of food quality control. 1. Microbiological analysis. Food and Drug Administration. FAO Food Nutr. Pap., 14 (4 Revis 1): 1-338.
- Carbone, D. & Faggio, C. (2016). Importance of prebiotics in aquaculture as immunostimulants: Effects on immune system of *Sparus aurata* and *Dicentrarchus labrax*. *Fish Shellfish Immunol.*, 54: 172-178. DOI: 10.1016/j.fsi.2016.04.011.
- Eldahshan, O.A. & Singab, A.-N.B. (2013). Carotenoids. *Pharmacog. Phytochem.*, 2 (1): 225-234.
- Forsatkar, M.N.; Nematollahi, M.A.; Rafiee, G.; Farahmand, H. & Martínez-Rodríguez, G. (2017). Effects of prebiotic mannan oligosaccharide on the growth, survival, and anxiety-like behaviors of zebrafish (*Danio rerio*). *J. Appl. Aquac.*, 29 (2): 183-196. DOI: 10.1080/10454438.2017.1306732.
- Gebreyohannes, G. & Gebreyohannes, M. (2013). Medicinal values of garlic: A review. *Int. J. Med. Med. Sci.*, 5 (9): 401-408.
- Gibson, G.R. & Roberfroid, M.B. (1995). Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *J. Nutr.*, 125: 1401-1412.
- Hoseinifar, S.H.; Mirvaghefi, A. & Merrifield, D.L. (2011). The effects of dietary inactive brewer's yeast *Saccharomyces cerevisiae* var. *ellipsoideus* on the growth, physiological responses and gut microbiota of juvenile beluga sturgeon (*Huso huso*). *Aquaculture*, 318 (1-2): 90-94.
- Hoseinifar, S.H.; Soleimani, N. & Ringø, E. (2014a). Effects of dietary fructo-oligosaccharide supplementation on the growth performance, haemato-immunological parameters, gut microbiota and stress resistance of common carp (*Cyprinus carpio*) fry. *Br. J. Nutr.*, 112 (8): 1296-1302.
- Hoseinifar, S.H.; Eshaghzadeh, H.; Vahabzadeh, H. & Mana, N.P. (2016). Modulation of growth performances, survival, digestive enzyme activities and intestinal microbiota in common carp (*Cyprinus carpio*)

- larvae using short chain fructooligosaccharide. *Aquac. Res.*, 47 (10): 3246-3253.
- Hoseinifar, S.H.; Sharifian, M.; Vesaghi, M.J.; Khalili, M. & Esteban, M.Á. (2014b). The effects of dietary xylooligosaccharide on mucosal parameters, intestinal microbiota and morphology and growth performance of Caspian white fish (*Rutilus frisii kutum*) fry. *Fish Shellfish Immunol.*, 39 (2): 231-236
- Mageney, V.; Neugart, S. & Albach, D.C. (2017). A guide to the variability of flavonoids in *Brassica oleracea*. *Molecules*, 22 (2) 252: DOI: 10.3390/molecules22020252.
- Marković, Z.; Dulic, Z.; Živić, I. & Mitrovic-Tutundzic, V. (2009). Influence of abiotic and biotic environmental factors on weight gain of cultured carp on a carp farm, *Arch. Biol. Sci.*, 61 (1): 113-121.
- Momeni-Moghaddam, P.; Keyvanshokoo, S.; Ziaei-Nejad, S.; Parviz Salati, A. & Pasha-Zanoosi, H. (2015). Effects of mannan oligosaccharide supplementation on growth, some immune responses and gut lactic acid bacteria of common carp (*Cyprinus Carpio*) fingerlings. *Vet. Res. Forum*, 6 (3): 239-244.
- Najim, S.M.; Al-Noor, S.S. & Jasim, B.M. (2014). Effects of fish meal replacement with fish biosilage on some haematological and biochemical parameters in common carp *Cyprinus carpio* fingerlings. *Int. J. Res. Fish. Aquac.*, 4 (3): 112-116.
- Ringø, E.; Dimitroglou, A.; Hoseinifar, S.H. & Davies, S.J. (2014). Prebiotics in finfish: An update. In: Merrifield, D. & Ringø, E. (eds.), *Aquaculture nutrition: Gut health, probiotics and prebiotics*. John Wiley & Sons, Oxford: 360-400.
- Ringø, E.; Sperstad, S.; Myklebust, R.; Mayhew, T.M. & Olsen, R.E. (2006). The effect of dietary inulin on aerobic bacteria associated with hindgut of Arctic charr (*Salvelinus alpinus* L.). *Aquac. Res.*, 37 (9): 891-897.
- Ringø, E.; Olsen, R.E.; Gifstad, T.Ø.; Dalmo, R.A.; Amlund, H.; Hemre, G.-I. & Bakke, A.M. (2010a). Prebiotics in aquaculture: A review. *Aquac. Nutr.*, 16 (2): 117-136.
- Ringø, E.; Løvmo, L.; Kristiansen, M.; Bakken, Y.; Salinas, I.; Myklebust, R.; Olsen, R.E. & Mayhew, T. (2010b). Lactic acid bacteria vs. pathogens in the gastrointestinal tract of fish: A review. *Aquac. Res.*, 41 (4): 451-467
- Taherpour, K.; Moravej, H.; Shivazad, M.; Adibmoradi, M. & Yakhchali, B. (2009). Effects of dietary probiotic, prebiotic and butyric acid glycerides on performance and serum composition in broiler chickens. *Afr. J. Biotechnol.*, 8 (10): 2329-2334.
- Wolf, K & Darlington, R.W. (1971). Channel catfish virus: A new herpes virus of ictalurid fish. *J. Virol.*, 8 (4): 525-533.



- Yang, J.-L. & Chen, H.-C. (2003). Serum metabolic enzyme activities and hepatocyte ultrastructure of common carp after Gallium exposure. *Zool. Stud.*, 42 (3): 455-461.
- Ye, J.-D.; Wang, K.; Li, F.-D. & Sun, Y.-Z. (2011). Single or combined effects of fructo- and mannan oligosaccharide supplements and *Bacillus clausii* on the growth, feed utilization, body composition, digestive enzyme activity, innate immune response and lipid metabolism of the Japanese flounder *Paralichthys olivaceus*. *Aquac. Nutr.*, 17 (4): 902-911.